



INTERNATIONAL JOURNAL OF PAIN MANAGEMENT

ISSN NO: 2688-5328

Research Article

DOI: 10.14302/issn.2688-5328.ijp-19-2731

Research Status and Application Prospect of Epigenetic Regulation of BDNF Gene Expression in Chronic Pain

Xiang Wei^{1,*}, Li Na-Wang¹

¹Hospital of Suzhou University, Suzhou, China

Abstract

The rise of epigenetics provides a new idea for studying the regulation of chronic pain-related genes and synaptic plasticity. External environmental stimuli can regulate BDNF genes through different epigenetic modifications. The epigenetic changes of the BDNF gene can affect the expression of its mRNA and protein and participate in the development of chronic pain. By reviewing the literature, this paper reviews the mechanism of epigenetic regulation of brain-derived neurotrophic factor (BDNF) in chronic pain, which provides some new directions and targets for the treatment of chronic pain.

Corresponding authors 15895573053@163.com	or Xiang	Wei,	Hospital	of	Suzhou	University,	Suzhou,	China,	Email:
Keywords: chronic pain; brain-derived neurotrophic factor (BDNF); Epigenetics; DNA methylation; Histone modification; Non-coding RNA regulation									
Received: Mar 25, 2019	А	ccepte	d: Jun 03,	2019	F	Published: Ju	un 13, 2019)	
Editor: Farah Yousef, Damascus University, Syria.									



Chronic Pain

Pain is an unpleasant subjective feeling and emotional experience associated with tissue damage or potential damage[1]. It is the fifth largest clinical sign in clinical practice and one of the most common clinical symptoms. Pain caused by peripheral tissue injury or disease Inappropriate treatment or treatment options and medications are inappropriate, Acute pain often develops into chronic pain(lasting more than 6 months).Chronic pain is a clinical syndrome characterized by pain, which is caused by a combination of biological, psychological and social factors. People with chronic pain are more than 20% in developed countries [2], and there are hundreds of millions of pain patients in China alone. Epidemiological surveys show that the prevalence of chronic pain in the elderly is 70 to 85%, and that of middle-aged people is 25 to 40%. With the aging of the population in China, pathological pain syndrome is more common among the elderly population, More than half of the population over 65 years old in China suffer from pain. Chronic pain lasts for a long time, stubborn and difficult to cure, therefore some negative emotional factors gradually breed, among which anxiety and depression occur. Epidemiological data show that 34 to 66 % of depression patients are accompanied by chronic pain, and up to 52% of patients with chronic pain have major depression [3]. Therefore, the search for effective treatment methods and drugs to improve the quality of life of patients and reduce the socio-economic burden is of great significance

The pathogenesis of chronic pain is complex. Previous research hotspots have focused on the mechanism of neuropathic pain from the perspective of molecular proteinology and signal transduction pathways. Now the genetic mechanism of neuropathic pain and the study of epigenetic mechanisms are receiving increasing attention. Numerous studies have shown that the production and maintenance of neuropathic pain is closely related to changes in the transcription and expression levels of pain-related molecules and receptor genes on primary sensory neurons, secondary sensory neurons, immune cells, and glial cells. How many pain-related genes are procedurally initiated, transcribed and translated, increased in expression, or specifically inhibited, decreased in expression, thereby regulating



pain, has now become a new direction of pain mechanism research, which involves the appearance of Genetics. The epigenetic changes are reversible, and the use of corresponding epigenetic interventions can re-express silent analgesic genes or reduce the expression of up-regulated pain-generating genes, providing new insights and Direction into the treatment of neuropathic pain.

Epigenetic Regulation

A large body of research evidence indicates that pain-related molecular changes after neurological damage are regulated by epigenetics. These genetic mechanisms are reversible modifications in the absence of changes in the DNA sequence. the heritability of the expression of the gene changes [4]. Epigenetic regulation refers to the structural adjustment of pretranscription genes at the chromatin level. It is a unique regulatory mechanism of the eukaryotic genome. Therefore, epigenetic regulation is also known as chromatin based gene expression regulation [5]. Epigenetics is the result of interaction between environmental factors and genetic material in cells. It is a dynamic bridge between environmental stimuli and genotypes. Environmental factors change the function of genes by acting on key enzymes. Epigenetic mechanisms have a lasting effect on genes that are activated in a neuralgia environment. Currently, DNA methylation, histone modification, and non-coding RNA regulation play a vital role in gene activation and inactivation as well as phenotypic metastasis[6, 7].

The formation of chronic pain requires a large number of genes to participate in it. Recent studies have reported that the gene expression of 12.4 % in chronic neuralgia rats is increased and 7 % is decreased [8]. The products of these genes can regulate the neural network adaptability required for chronic pain production in a long-term and stable manner, and continuously and stably change gene expression and heritability, which are the two most important features of epigenetic regulation. Thus, changes in gene transcription activity in epigenetic regulation mediate the production and maintenance of chronic pain. Therefore, the rise of epigenetics provides a new idea for the study of chronic pain-related gene regulation [9]. At present, epigenetic modifications that have received attention in the field of pain and analgesia



Pen Occess Pub

mainly include: DNA methylation, non-coding RNA regulation, and histone modification, etc. They regulate pain-related molecular, Synaptic, and nuclear network activities in nervous system, producing different painful behavior phenotypes, including neuralgia, somatic pain, and visceral pain. (Figure 1)

Brain Derived Neurotrophic Factor

Brain derived neurotrophic factor (BDNF) is another vital neurotrophic factor discovered after nerve growth factor (NGF). It can maintain the normal physiological functions in the central and peripheral nervous system, plays an important role in the survival, differentiation, development of neurons, and can promote the regeneration of injured neurons[10]. With the deepening of research, BDNF, as a signal molecule connecting neuronal-glial intercellular information transmission, has attracted more and more attention in the development of pain [11]. In recent years ,A large number of studies have found that epigenetic modifications play an important role in the expression of BDNF gene. In the central system, the BDNF gene is the first recognized downstream target gene under epigenetic regulation. These studies provide a new direction for the pathogenesis of chronic pain and provide a new target for drug therapy. This article reviews the study on epigenetic regulation of BDNF gene and chronic pain.

BDNF Gene and Chronic Pain

Structure of BDNF Gene

BDNF has a complex structure, which has been studied in depth[12, 13]. Many studies have shown that rodents and humans have substantial similarities. The rat BDNF gene contains 9 exons, Which consists of 8 non-coding exons and 1 exon encoding the protein. Any of the 8 non-coding exons can be selectively spliced with the coding exon and transcribed to form BDNF mRNA. The human BDNF gene is located on chromosome 11 and encodes 247 amino acids. The protein contains 11 exons, each of which can be independently cleaved into a coding sequence, in which 9 promoters are present on exons I, VII, VIII. And IX, all of which initiate the transcription process. The coding sequence forms a different BDNF transcript[14] The study found that external environmental stimuli can regulate different promoters of BDNF through different epigenetic modifications, which can cause changes in mRNA expression of different exons

and ultimately lead to changes in BDNF gene expression.

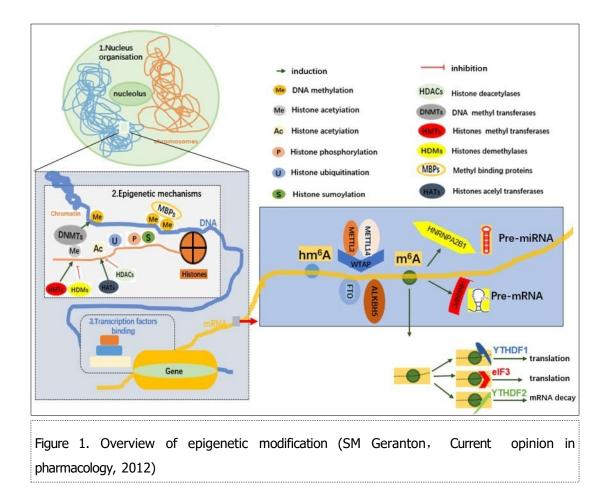
Human and rodent BDNF genes are contains nine exons (Figure 2), Which has their own promoter, producing a variety of different transcripts. Interestingly, all of their transcripts were translated into the same BDNF protein[12, 13, 15, 16]. Some exons of BDNF between humans and rodents are highly conserved [17]. Each exon is regulated by its own unique promoter for the temporal and spatial specificity of BDNF expression in an activity-dependent manner.

BDNF gene is Involved in the Development of Chronic Pain

Multiple patient and animal studies of chronic pain have shown that abnormal BDNF gene expression may be involved in the pathogenesis of chronic pain. Numerous studies have shown that BDNF is an important substance regulating nociceptive information, and participating in the occurrence and maintenance of pain sensitization [19]. As early as 1999, the researchers found that increasing the concentration of BDNF in the brain increased the sensitivity of rats to pain, and the use of BDNF inhibitors significantly reduced the pain performance in rats[20].In the CPIP model induced by rodent hind paw ischemia-reperfusion (IR) injury, microglia accumulate in the ipsilateral dorsal horn after ischemia for 3h, and the accumulated microglia release BDNF, increasing the excitability of neurons in the dorsal horn. And produce painful behavior [21]. Studies have also found that spinal nerve ligation can increase the activity and number of neurons producing BDNF, and then the concentration of BDNF increases. This result indicates that the damage will increase the concentration and activity of endogenous BDNF [22]. Other studies have also demonstrated this conclusion, such as temperature - induced hypersensitivity associated with elevated BDNF levels [23]. The intensity of pain in chronic pancreatitis is directly proportional to the concentration of BDNF [24]. These results indicate that pain does increase the expression of BDNF. During nociceptive stimulation in chronic pain, nociceptors are activated and release BDNF [25]. The study found that the expression of BDNF was significantly up-regulated in the posterior horn of spinal cord in the pain-sensing (somatic pain) caused by cutting and the pain model (neural pain) caused by lumbar 5 root excision [26]. In an animal model of neuropathic pain induced by nerve injury, increased







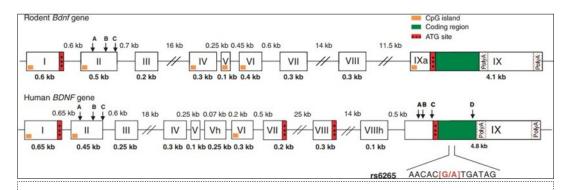


Figure 2. Structure of human and rodent BDNF genes. The exon is a box and the intron is a line. The sequence number of the exon is represented as a Roman numeral, and the molecular weights of the exons and introns are represented as Arabic numerals. Two polyadenylation sites (poly A) are encoded exons. The start codon ATG of the marker transcription initiation is indicated by a red box. The exon IX region encoding the pro-BDNF protein is shown in green boxes.. There are also some exons, such as exons II and IX, with alternative splice donor sites (A, B, C, D) containing different transcript variants.[18]



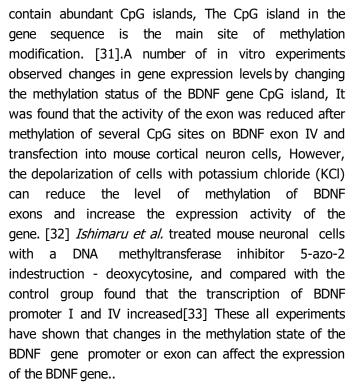
expression of BDNF can be observed in intermediate cell neurons, large cell neurons, and their central terminals and spinal cord of dorsal root ganglia [27] intrathecal administration. BDNF scavenger TrkB/Fc can neutralize endogenous BDNF, thereby blocking its binding to receptors, and alleviating hyperalgesia induced by nerve injury [28]. Geng et al found that early spinal cord dorsal rat spinal nerve ligation model The concentration of BDNF is significantly increased, combined with changes in the pain threshold of the rat, they believe that BDNF in the spinal dorsal horn is associated with the onset of neuropathic pain [29]. Coull et al found that injection of BDNF into the spinal cord of normal mice induced symptoms of neuropathic pain in mice, whereas inhibition of BDNF mRNA expression or inhibition of BDNF and receptor by antisense RNA was first used. The combination and destruction of BDNF signaling process, peripheral nerve injury no longer induces neuropathic pain, this result clearly shows that BDNF takes part in the occurrence of neuropathic pain, its role is blocked will inhibit pain Occurs, and subsequent experiments further demonstrate the mechanism of action of BDNF in the pathogenesis of neuropathic pain [30]. The above studies suggest that BDNF plays a key role in the generation and development of neuropathic pain.

BDNF is important for the regulation of pain, and epigenetics participates in the occurrence and development of pain. Recent research has focused on the epigenetic mechanism of the BDNF gene, which affects the BDNF gene by regulating environmental factors. Therefore, the lasting expression changes contribute to the production and development of neurological pain. The epigenetic changes of the BDNF gene can affect the expression of its mRNA and protein and participate in the development of chronic pain. However, there are few studies on how epigenetic modification regulates the expression of BDNF genes in pain. The following will be discussed further.

Regulation of Methylation on Expression of BDNF Gene and Chronic Pain

Histone and DNA Methylation

The methylation modification of BDNF gene has an important regulatory effect on the expression of this gene. *Boulle et al.'s* sequence analysis of the BDNF gene shows that human and rat BDNF gene promoter I, II, IV



In-vivo experiments also showed that the methylation state of the gene is related to the expression of the BDNF gene. Studies have shown that repeated morphine injection may induce demethylation of two CpG sites in 197 and 176 of the BDNF exon IV gene promoter, which increases the expression of BDNF mRNA and protein levels in DRG neurons Moreover.

Intrathecal injection of anti-BDNF antibodies can inhibit mechanical allergic reactions in rats after repeated morphine exposure. By increasing BDNF in DRG neurons, Inhibition of DNA methylation can significantly promote the production of opioid-induced hypersensitivity reactions. The results show that the methylation level of the BDNF gene exon in DRG neurons can regulate the expression of the BDNF, which is of great significance in opioid-induced hypersensitivity reactions. [34]. Ikegame and other studies found that BDNF methylation modification not only existed in brain tissue. It also exists in peripheral blood cells and saliva [35]. Several studies have found changes in DNA methylation levels of the BDNF gene in the brain and peripheral blood of patients with chronic pain. A clinical study using blood to isolate genomic DNA found that women with myalgia syndrome showed significantly abnormal DNA methylation compared to age-appropriate health controls. Muscualgia syndrome is associated with different methylation of multiple genes, such as BDNF,





HDAC4, etc.. [36].These studies have caused changes in the methylation level of animal BDNF gene by changing physical, chemical, and other environmental factors, thereby affecting the expression of BDNF gene The study found that there was a change in the level of total DNA methylation and related enzymes and binding proteins during the pain process. Expression changes of histone demethylase JMJD3, JMJD2A, transcription Suppressor MeCP2, histone dimethyltransferase G9a and DNA methylase DNMT in pain.

JMJD3

Recent studies have found that specific demethylation H3K27me2/3 produces deinhibitory histone demethylase JMJD3 also appears to be involved in the pain mechanism. It has been found that BDNF increases after peripheral nerve injury, leading to the occurrence of neuropathic pain. Neuropathic pain. Intrathecal administration of anti-BDNF antibodies inhibits thermal hyperalgesia mechanical and allodynia [37].JMJD3 can regulate the expression of BDNF in DRG neurons after nerve injury Generally, PRC2 silences the BDNF gene, and PRC2 contains the catalytic subbase EZH2. After stimulating neurons in vitro using NMDA, JMJD3 was recruited into the BDNF promoter to induce BDNF gene promoter demethylation .Therefore, the deinhibition of the promoter helps to increase the BDNF expression in mature neurons[38].In addition to promoter inhibition, BDNF is also enhanced through the acetylation mechanism and through the role of CREB kinase / CBP[39]. In addition to its contribution to the pain process, JMJD3 also participates in the inflammation mechanism. JMJD3 increases after inflammation such as LPS.Studies have also been found that transcriptional regulation of Stat1 and Stat3 activates the expression of inflammation related genes in macrophages cultures of microglia. For example, Lee et al. described an increase in spinal cord injury (SCI) produced by JMJD3 in endothelial cells, which result in the elevation of cytokine IL-6 via demethylation of its promoter[40]. The incident has been confirmed in vitroand JMJD3 siRNA inhibits IL-6 expression in oxygen/glucose deprivation reactions. Therefore, the epigenetic modification of the BDNF promoter plays a vital role in the activation of neurons, and an increase in the level of BDNF may be observed after the injury of neurons. JMJD3 may be a Potential target for reducing neuropathic pain after neurotraumatic lesions by reducing inflammation and neurotrophic factor expression.

JMJD2A

JMJD2A, also known as KDM4A or JHDM3A, is a histone demethylase. It can catalyze the demethylation of H3K9 and H3K36 in the trimethylation / dimethylation state to different methylation levels(H3K9me2/3 and H3K36me2/3). In addition, JMJD2A can also act as a coactivator or cosuppressor binding protein of a transcription factor to regulate gene expression at the transcription level. The study found that the VPA treatment of rat embryonic neural stem cells led to a strong increase in the expression of BDNF mRNA, which was related to the demethylation of H3K9 at the Bdnf IV promoter in rats. It was further found that both KDM2A and KDM2C occupied the exons 5 of the rat promoter IV and BDNF genes under VPA conditions. At the same time, the reduction of the VPA-mediated H3K9me3 level was reduced by siKDM4A/C, while the level of H3K36me3 in the BDNF promoter did not change significantly. These results show that KDM2A/C is necessary for the increase of BDNF expression during neuron differentiation and is related to the demethylation of the promoter region H3K9me3. [41]. In addition to participating in neuronal differentiation, spinal cord JMJD2A dependent histone demethylation also plays an important role in traumatic hypersensitivity reactions. Zhou et al. found that the mRNA and protein of JMJD2A were increased highly in neuronal neurological pain induced by CCI or SNI of the sciatic nerve. The JMJD2A participates in and maintains the development of neurotic pain by combining with the BDNF promoter and demethylated H3K9me3 and H3K36me3 on its promoter to upgrade the expression of BDNF. In addition, the knockout of JMJD2A blocked the hypersensitivity response of mouse with chronic pain caused by CCI and SNI. These results show that the increase of JMJD2A promotes the neurological pain of BDNF, which can treat as a hopeful target for the cure of chronic pain. [42].

MECP2

The study found that there was a change in the level of total DNA methylation and related enzymes and binding proteins during the pain process. MeCP2, or methylated CpG binding protein, is the most clearly studied member of the currently methylated DNA binding



protein(DNA-binding protein, MBD) family [43]. In the nervous system, the BDNF gene is the first recognized downstream target gene regulated by MeCP2 [44]. In recent years, more and more evidence has shown that MeCP2 is closely related to the formation of chronic pain. MeCP2 binds to the CpG site of methylation on DNA to inhibit the transcription of specific genes, while MeCP2 after phosphorylation loses its binding ability. [45]. The study found that the total level of DNA methylation and methylation of MeCP2 mRNA levels in the neurological pain model induced by chronic sciatic compression injury (CCI) were significantly increased. After the intervention of 5-azacytidine(5-azacytidine, 5-AZA), the total DNA methylation level and MeCP2 levels were reduced. At the same time, CCI induced mechanical hyperalgesia thermal hyperalgesia and were significantly relieved [46]. MeCP2 is a protein related to Rett syndrome. Clinical studies have found that patients with Rett syndrome with MeCP2 mutation show insensitivity to pain. The binding pattern of MeCP2 on the genome also changed after nerve injury. The study found that MeCP2 expression in the central amygdala (CeA) increased and MeCP2 local silencing alleviated inflammatory pain in the state of sustained inflammatory pain, while the over-expressed MeCP2 mice showed pain sensitivity and a large number of gene expression changes such as BDNF in the central amygdala [47]. Some studies have shown that the formation of polymers between MeCP2 and HDACs has an inhibitory effect on the expression of BDNF promoter IV. After depolarization of rat nerve cells in vitro experiments, it was found that the phosphorylation level of MeCP2 increased. However, increased phosphorylation of MeCP 2 can separate the binding sites of MeCP 2 and BDNF gene, thus relieving the inhibition of expression of BDNF gene. [48].In addition, DNA methylation in the prefrontal cortex and amygdala decreased after nerve damage, and this decrease is closely related to the degree of pain [49]. This suggests that nerve injury induced DNA methylation in the central nervous system has mediated the development of pain.

In the nucleus accumbens, chronic stress increases the DNA differentiation of the neurotrophic factor (gdnf) gene promoter source and the binding of MeCP2. MeCP2 reduces the acetylation of the histone H3 on the gdnf promoter through interaction with HDAC2., This reduces gdnf gene expression and produces anxious



behavior [50]. Therefore, the relationship between DNA methylation and BDNF in chronic pain remains to be further explored.

Histone Dimethyl Transferase G9a

G9a(encoded by Ehmt2) is an H3K9 methyltransferase that belongs to a typical histone methyltransferase and is responsible for gene silencing. Recent studies have found that G9a is also closely related to pain formation. As mentioned above, methylated CpG binds methyl-CpG binding proteins, such as MeCP2. After MeCP2 binds to methylated DNA, it will raise transcription factor formation complexes such as G9a [51]. Therefore, G9a may be an essential epigenetic marker protein for MeCP2 inhibition of gene transcription in chronic pain

In fact, a large number of pain-related genes, such as BDNF, are subject to the epigenetic regulation of G9a in many environments[47].Zhang et al. found that the expression of G9a and H3K9me2 proteins in CeA central amygdala in mice with persistent inflammatory pain was reduced. The level of G9a on the BDNF gene promoter was also reduced and accompanied by an increase in BDNF protein levels. CeA local injection of G9a inhibitors reduced H3K9me2 and increased BDNF protein levels, and delayed the recovery time of persistent inflammatory pain [47]. In addition, the pain threshold in mice with G9a transgenic, CeA local injection virus silent G9a expression, decreased.

A recent research showed that Mecp2 expression increased in the central amygdala (CeA) of mice due to CFA-induced chronic inflammatory pain and repeated morphine exposure. The increased MeCP2 binding and inhibition of transcription inhibition factor factor protein methyltransferase G9a resulted in a decrease in the inhibition marker H3K9me2 catalyzed by G9a in CeA, and inhibition of G9a increased the expression of BDNF.

DNMT

Recent studies have related DNA methylation of specific genes with pain allergic reactions. Current research has found that SNL increases the binding of DNMTs to BDNF promoter, which means SNL increases the spinal cord DNA methylation of BDNF promoter in rats. It is worth noting that although SNL increases the combination of DNMTs with BDNF promoters. At the same time, Tet1 dependent DNA demethylation of spinal cord was found to take part in the process of injury perception. Tet1 mediates the conversion of 5-methylcytosine(5mC) to



5- hydroxymethylcytosine(5hmC), thereby promoting DNA demethylation. In the SNL rats model, the Tet1 expression was increased in the dorsal angular neurons and the CpG dinucleotide hydroxylation in the BDNF promoter was 5mC to 5hmC to promote the expression of the spinal cord BDNF after surgery. The concentration of Tet1 binding and 5hmC was reduced by focal knock of Tet1 expression in spinal cord. The concentration of 5mC in CpG site in BDNF promoter was further increased and the expression of BDNF in spinal cord was reduced, while the abnormal pain was reduced [52].

In addition, SNL upgrade Tet1 expression also inhibits the binding of DNA methyltransferase(DNMTs) to the BDNF promoter, which is a catalyst. The necessary conditions for the transcription silence of metron-5-cytosine (5C) to 5mC. In summary, these data show that at the CpG site of the BDNF promoter, SNL increased Tet1 expression promotes DNA demethylation by converting 5mC to 5hmC and inhibiting DNMT binding to regulate spinal cord BDNF expression, thus leading to the development of abnormal behavioral pain. (Figure 3 and Table 1).

Regulation of Histone Acetylation on Expression of BDNF Gene and Chronic Pain

Histone modification as an important epigenetic regulation also affects BDNF gene expression. Histone modifications include methylation, acetylation, phosphorylation and many other regulatory modes. Histone modifications can affect the binding of genes through the regulation of histone structure, leading to changes in gene expression. Some studies have shown that histone tails (methylation of H3 and H4), Modifications, acetylationwhich cause transcription of inflammatory molecules, such as cytokines and chemokines, are responsible for the development of chronic pain. In these cases, HAT appears to be involved in chemokine expression, and HDAC is associated with cytokine expression, which together lead to neuropathic pain.Next we mainly discuss the effect of histone acetylation on the BDNF gene.

In the chronic neuralgia model, acetylation occurs on many pain-related gene promoters of DRG, such as opioid receptors and bdnf genes [53]. The increase of H3 and H4 acetylation on the BDNF gene promoter induced by nerve damage and inflammation, the expression of the BDNF gene was increased, and excitatory and inhibitory



Synaptic transmission was involved, causing the production and maintenance of neuralgia. In the persistent inflammatory state, the expression of BDNF regulated by histone acetylation is increased, and chronic pain behavior is produced by activating TrkB and initiating Synaptic metastasis in the middle slit macronucleus (Glualyaji) via the PLC-PKC signal pathway[54, 55]. Neural injury promotes the increase of histone H3 and H4 acetylation in the BDNF promoter region in DRG and spinal cord injury to regulate the expression of BDNF, and can help induce or maintain nerve pain [56, 57]

Ishimaru et al. treated mouse neuroblastoma cells with the HDACs inhibitor trichostatin, and discovered that the acetylation levels of histone H3 and H4 bound to BDNF promoter I were increased, while the expression of BDNF gene was also increased [58]. Subsequent animal experiments have also dig out that elevated standard of histone acetylation are positively correlated with expression of BDNF [59, 60]. Uchida H et al. showed that the initial increased expression of BDNF exon I controlled by epigenetic mechanisms may join in the development of neuropathic pain. In a partial sciatic nerve ligation model, they found that histone H3 and H4 acetylation regulate BDNF promoter transcription. Chromatin immunoprecipitation (ChIP) assay showed that genomic H3 H4 acetylation of BDNF promoter I increased and significantly on one day after nerve injury, and histone acetylation levels remained elevated for at least 7 days. Epigenetic control of protein acetylation is required for the induction and maintenance of BDNF promoter transcription in DRG [61]. Similarly, studies have shown that mechanical hyperalgesia and elevated expression of BDNF/SYN1 in the spinal dorsal horn can be induced by inoculation of Walkers256 tumor cells in rat tibia. Intraperitoneal injection of the HDAC inhibitor, suberoylanilide hydroxamic acid (SAHA), reverses these changes and exhibits an antinociceptive effect. These findings suggest that regulation of histone acetylation levels is at least partially involved in BDNF-induced synaptic plasticity in an indirect manner, thereby inducing pain central sensitization in BCP rats.

Winston et al. [62] gave intermittent chronic stress stimulation to pregnant rats at a certain intensity, and found that female offspring presented strong and sustained intestinal hypersensitivity response to transient stress stimulation of this intensity as adults, which was





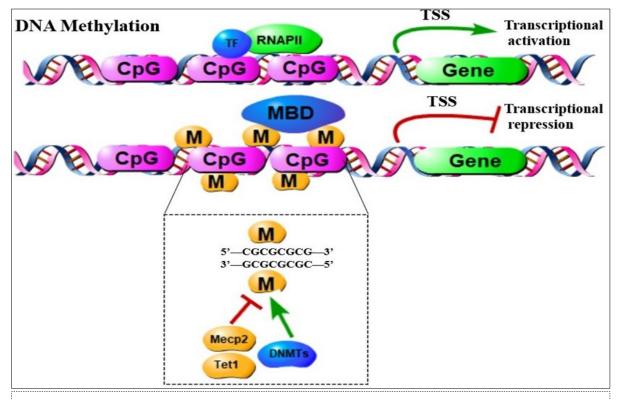


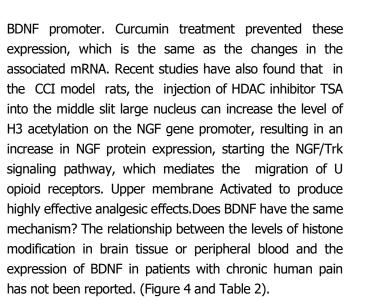
Figure 3. When DNA methyltransferase (DNMTS) catalyzes the addition of cytosine at the 5' position to the methyl group, DNA methylation happened. When there is no DNA methylation, the transcription factor and RNA polymerase 2 (RNAP II) bind to DNA, leading to gene expression. Recent researches suggests that members of the TET family are likely to undergo active DNA demethylation. DNA methylation cause transcriptional repression by recruiting proteins with a methyl binding domain such as MeCP2, which can further recruit a co-inhibitor complex containing HDAC. TSS, transcription start site

Table 1. I	Table 1. Epigenetic enzymes related to neuropathic pain							
Enzyme	Alteration	Molecular effect	Effect on gene expression	Gene altered	Direct relation to pain	Reference		
JIVLJD3	increased	Demethylation of H3k27me2/3	De-repression	BDNF,IL-6	NO	Uchida, H et a1,2013; ³⁷ Ernest, P et a1,2016 ³⁸ , ³⁹		
JMJD2A	increased	H3k9me3/ H3k36me3	Gene expression	BDNF,5-HT	NO	Zhou, J et a1,2017 ⁴²		
MECP2	increased	Binding to CpG	Gene silencing	BDICF	YES	Wang, Y et a1,2011 ⁴⁶ Zhang, Z et a1,2014 ⁴⁷		
DNMT	increased	Binding to CpG	Gene expression	BDNF	YES	Hsieh, M et a1,2016 ⁵²		
Tetl	increased	Demethylation	Gene expression	BDNF, 5inc-51unc	YES	Hsieh, M et a1,2016 ⁵²		
G9a	decreased	H3k9ine2	Gene silencing	BDNF	YES	Zhang, Z, et a1,2014 ⁴⁷ Ooi, L et a1,2007 ⁵¹		



stronger than male offspring. However, blocking BDNF protein receptor or knocking BDNF expression by small interfering RNA technology can reduce the intestinal hypersensitivity of female offspring, indicating that it is related to up-regulation of BDNF protein level in spinal dorsal horn. In addition, continuous administration of HAT inhibitor curcumin in female offspring can alleviate the stress related hypersensitivity of the newborn, while the mRNA and protein level of BDNF decreased, suggesting that HAT inhibitor may alleviate the intestinal hypersensitivity caused by chronic stress by inhibiting the expression of BDNF.

Curcumin has long been recognized as an inhibitor of HAT activity and used to treat of neuropathic pain. Studies have found that curcumin has an antinociceptive effect in a neuropathic pain model of chronic compressive injury (CCI) rats. The effects of curcumin on P300/CBP and BDNF was studied in a model of chronic compression injury. Treatment with curcumin for 7 consecutive days significantly attenuated CCI- induced thermal hyperalgesia and mechanical allodynia. Chromatin immunoprecipitation analysis showed that curcumin can reduce the recruitment of p300/CBP and acetyl histone H3/acetyl histone H4 to BDNF gene promoters. A similar result was also observed after curcumin treatment. These results indicate that curcumin have a therapeutic action in neuropathic pain by weakened BDNE gene expression mediated by p300/CBP HAT activity.[63] Another group studied the neurotic pain induced by HAT activity in rat CCI associated with transcription coactivator P300. They invented that the expression of P300 in the spinal cord was increased after and that intrathecal injection of the slow virus CCI, expressing P300 shRNA reduced the increase of P300 and reduced the increase of BDNF mRNA in the spinal cord. All the time, these reductions are followed with reductions in the hypersensitivity reactions caused by CCI. Intrathecal administration of the HAT inhibitor C646 reproduces the results. they further studied the p300/CBP, which stand for two functionally interchangeable proteins, expressed BDNF and COX2 in the spinal cord. Both the P300 and CBP are copositioned with BDNF or COX2 in the dorsal neurons of the spinal cord. CCI animals are known to have BDNF and COX2 increases in the spinal cord. Consistent with these facts, nerve damage causes P300, CBP, and H3K9ac, but not the increase in the binding of H4K5ac to the BDNF promoter, and all these proteins are combined with the



MiRNA Regulation of BDNF Gene Expression and Chronic Pain

Neurological pain is chronic pain caused by somatosensory system diseases. MicroRNAs (miRNAs) have been extensively studied for the development of neurological pain and neuroinflammation caused by nerve damage. But, the specific mechanisms by which miRNAs are involved in neuropathic pain remains unknown.

The target miRNA of the BDNF gene can directly target the 3-UTR region of its target gene mRNA. The 3-end non-translation region(3 '-UTR) of the BDNF is an important binding region of the miRNA and recognizes the target mRNA through base complementary pairing., According to the different degree of complementarity, the silent complex degrades the mRNA of the target gene or deters the translation of the target gene, resulting in a change in the level of expression of the target gene. Multiple miRNAs have a regulatory effect on the BDNF gene. Studies have shown that the abnormal regulation of microRNA on BDNF is related to chronic pain. Genda et al. found that 111 types of miRNA expression were significantly regulated in CCI rats in the spinal cord dorsal after surgery [64]. The study found that miRNA-195 inhibited the expression of BDNF gene [65]. In the rat model of SNI, the expression of miRNA-195 in the DRG was increased [66], After the spinal nerve ligation, the expression of miRNA-195 in the spinal cord was increased and continued to be at least 14d [67]. Up - regulation of miRNA-195 expression strengthened mechanical pain and pain allergies after peripheral nerve damage, and the opposite result was observed after the use of miRNA-195







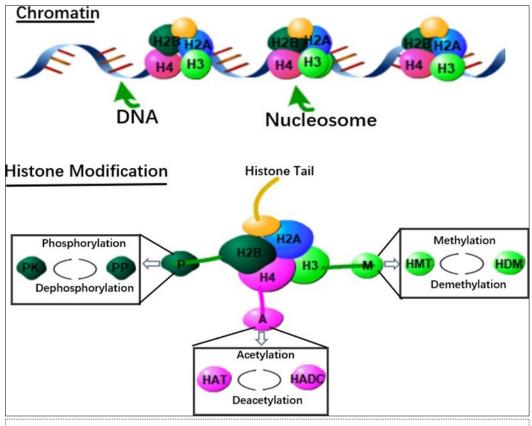


Figure 4. In the chromosome, DNA is wrapped around a single histone to form the basic unit of DNA packaging - the nucleosome. Epigenetic modification on histones for histone translational histone acetyltransferase (HAT; acetylated histone), histone deacetylase (HDAC; removal of histone acetyl), histone methyltransfer Enzyme (HMT; methylated histone), histone demethylase (HDM; removal of histone methyl), enzymatic mechanism of protein kinase (PK; phosphorylated histone) and protein phosphatase (PP; removal) Histone phosphate group).

Table 2. Histone acetylation related to neuropathic pain							
Pain model	Enzyme	Inhibitor	Histone acetylation	Site	Gene altered	Reference	
Visceral pain	HAT	Curcumin		SP	BDNF	Winston, et a1,2014[62]	
Neuralgia Pain	/		H31114	DRG	BDNF	Uchida, et a1,2013; ⁶¹	
Inflammatory pain	/	/	H31114	Central nucleus	BDNF	Uchida,et a1,2013; ⁵³	
SNL	/	/	H31114	DRG	BDNF	Wang x, et al.2014	
BCIP	lidac	SAHA		DRG	BDNF	Hoget a1,2016	
CCI	HAT	Curcumin	H31114	SP	P300/CBP/BDNF/ COX-2	Zhu, X., et al.2014 ⁶³	
CCI	HAT	C646	НЗК9ас	SP	P300/CBP/BDNF/COX-2	Zhu, I, et al.2014	
CCI	HDAC	TSA	113	Central nucleus	NGF	Tao, W., et a1.2016	



inhibitors [67]. It is speculated that miRNA-195 may participate in the development of chronic pain by regulating the expression of BDNF.

Studies have shown that miRNA-132 can indirectly affect the secretion of BDNF by regulating MeCP2 in the hippocampus of rats [68] Similarly, in the formalin induced acute pain rat model, miRNA-124a was lowered in the spinal cord, which was related to the increase of MeCP2 mRNA. [69] Intravenous administration of miRNA-124a inhibitors aggravated pain behavior, and miRNA-124a mimics significantly attenuated formalin-induced inflammatory pain. [69] In another study, intrathecal injection of miRNA-124 was used to treat persistent pain allergies induced by reversible corner fork gum and to prevent the occurrence of mechanical abnormal pain in the SNI model rats [70], Possible mechanism is to alleviate neuropathic pain by inhibiting the expression of BDNF mRNA and protein

More data on the effects of analgesic and pain-inducing miRNAs. miRNA-183 achieves analgesia by targeting the regulation of Nav1.3 and BDNF expression in DRG.

Intrathecal injection of lentivirus expressing miRNA-183 restored SNI-induced down - regulation of miRNA-183, resulting in decreased expression of Nav1.3 and BDNF mRNA in DRG and significantly attenuated SNL-induced mechanical allodynia in rats. [71] Neuropathic pain in rats CCI leads to down-regulation of miRNA-1 in the sciatic nerve. After CCI, miRNA-1-targeted pain-related proteins BDNF and Cx43 are up- regulated in the sciatic nerve and DRG, ipsilateral to the rat. Significant mechanical allodynia occurred in the hind paws [72]. Lin et al found that the expression of miRNA- 183 in the dorsal root ganglia was significantly down-regulated after L5 spinal nerve ligation, and this was associated with the maintenance of mechanical allodynia. Furthermore, intrathecal injection of a lentivirus expressing miRNA-183 significantly attenuated mechanical allodynia and was associated with a significant downregulation of BDNF [73]

In addition, CCI-induced rat hypersensitivity was reduced by the use of miRNA- 206 mimic, a synthetic double-stranded RNA mimicking endogenous miRNA-206, and miR-206 in rat dorsal root ganglia after CCI was down-regulated, while the expression of BDNF mRNA and protein was increased in vivo. miRNA-206 mimetic



attenuates mechanical allodynia and thermal hyperalgesia by inhibiting the expression of BDNF mRNA and protein in CCI rats. These results indicate that miRNA-206 attenuates the development of neuropathic pain by targeting BDNF, indicating miRNA- 206 may be a potential target for the treatment of neuropathic pain [74].

The study by Li Haigin recorded for the first time the role of miRNA-375 / JAK2/STAT 3/BDNF signal conduction in the development of chronic morphine tolerance. Studies have shown that miRNA-375 can negatively regulate JAK2 to inhibit BDNF expression, thereby blocking the development of tolerance. The miRNA -375 level in the morphine tolerance model is reduced, and the JAK2 expression in chronic morphine therapy is negatively related. Lowering miRNA-375 levels can induce BDNF expression and induce behavior similar to pain allergies and spinal cord neuronal sensitivity, and ectopic expression of miRNA-375 or JAK2 participates in chronic morphine tolerance. The upward adjustment of miRNA-375 improved the morphine tolerance in the dorsal root ganglion of mice by inhibiting the JAK2/STAT 3 pathway, and the upward adjustment of miRNA-375 could inhibit the expression of BDNF by lowering the JAK2 in mice that tolerate morphine[75]. The study found that the miRNA-219 was reduced after the CFA injection in rats and the CpG island on the gene promoter was demethylated, and CaMKII y was involved in CFA-induced pain by regulating the dorsal horn of the spinal cord. Studies have shown that miRNA-219 promotes the resistance to injury in mice DRG, followed by continuous morphine treatment. After chronic morphine treatment, the expression of miRNA-219 in the dorsal root ganglion of mice was reduced, and the expression of CaMKII y was increased. The expression of miRNA-219 and CaMKII y are closely related to the development of morphine tolerance. The expression of proteins and mRNA in BDNF in the DRG is induced by time-dependent methods through prolonged morphine exposure, which is subject to the transcriptional regulation of miR-219 and CaMKII y. Removal of the DNF TrkB-Fc can partially reduce morphine tolerance. Overexpression of miRNA-219 using miRNA-219 simulators or the reduction of CaMK1I y expression using CaMK1I y small interference RNA significantly delayed morphine tolerance. In addition, hyperalgesia and spinal cord neuron sensitization were caused by knockout miRNA-219 in control mice or functional inhibition of miRNA- 219, both of





which were inhibited by CaMK1I γ small interference RNA or TrkB-Fc.[76] These results prove that miRNA-219 promotes the development of chronic tolerance of morphine analgesia in the DRG of mice by targeting CaMKII γ and lift the expression of CaMKII γ dependent BDNF.

Alternatively, beneficial effects can be achieved by inhibiting pain-induced miRNAs. In the SNI model, the expression of miRNA-195 was up-regulated in DRG [77]; the expression of miRNA-195 was up-regulated for at least 14 days after spinal nerve ligation. Up-regulation of miRNA-195 expression potentiated mechanical pain and hyperalgesia following peripheral nerve injury, whereas the opposite was observed with miRNA-195 inhibitors [78]. The study found that miRNA-195 can regulate the expression of BDNF gene [79]. It is speculated that miRNA-195 may participate in the development of chronic pain by regulating the expression of BDNF. In bone cancer induced pain model mice, it can induce an upward adjustment of p-CREB and CRTC1.

Adenovirus expression of CRTC1-siRNA by intrathecal injection effectively relieved mechanical pain and spontaneous pain caused by bone cancer by central inhibition of CRTC1, parallel to inhibition expressed by spinal cord BDNF and miRNA-212 / 132. Since the activation of the signal and the formation of a positive feedback loop of its target gene contribute to neuronal plasticity, an increase in CRTC1 can enhance the transcription of the CREB gene in the spinal cord and contribute to the maintenance of chronic pain. CRTCI-SiRNA weakens pain behavior induced by bone cancer and reduces the expression of CREB/CRTCI-target genes in the spinal cord, including BDNF and miRNA-212/132. [80] Analgesia is also produced by inhibition of miRNA- 221, which is caused by up-regulation of cytokine signaling inhibitor 1, a protein that inhibits cytokine signaling. After SNI, miRNA-221 is up-regulated in spinal microglia. Spinal cord miRNA-221 inhibitor down-regulates pro-inflammatory cytokines and reduces neuropathy-induced hypersensitivity [81] (Figure 5 and Table 3).

Summary and Expectations

BDNF has a wide and important role in the function of the central nervous system. The study of the effect of epigenetic modification on the expression of BDNF genes has attracted more and more attention. As an important molecular control mechanism in organisms, epigenetic regulation closely links complex genetic factors with diverse environmental changes. In both animal and clinical studies, it was found that the regulation of BDNF gene by epigenetics was related to chronic pain.

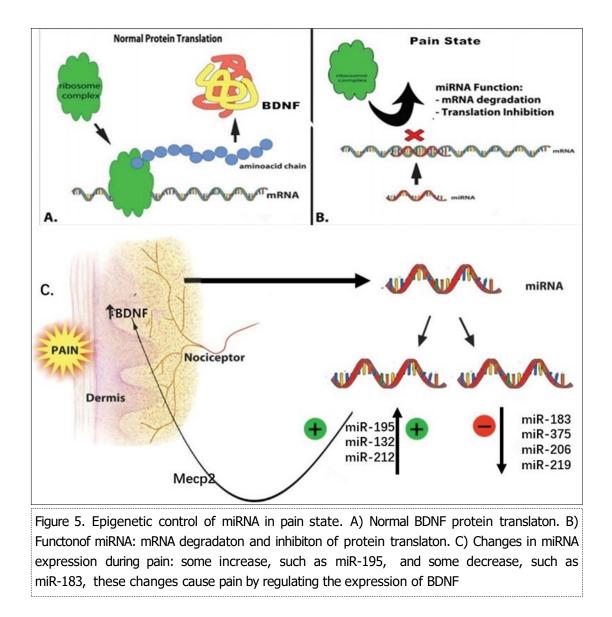
Based on the literature of recent years, the epigenetic mechanism of BDNF gene regulation for chronic pain needs to be further studied. It mainly includes the following aspects:

(1) To clarify the effects of pain on the expression of BDNF genes in different brain regions, That is, under the pain state, the expression of BDNF gene is raised or decreased in a certain brain area, and the effect and

Table 3. miRN/	Table 3. miRNAs related to neuropathic pain.						
miRNA	Alteration	Site	Gene altered	Reference			
miR-206	decreased	DRG	BDNF	Sun, et a1,2017 ⁷⁴			
miR-1	decreased	Sciatic nerve	BDNF/Cx43	Neumann, et a1,2015 ⁷²			
miR-183	decreased	DRG	BDNF/Nav1.3	Lin, et a1,2014 ⁷³			
miR-375	decreased	DRG	BDNF/STAT3/JAK2	Li, et a1,2017 ⁷⁵			
miR-219	decreased	DRG	BDNF/CaMICIIT	Hu,et a1,2016 ⁷⁶			
miR-212/132	increased	SP	BUNT CRTC' 'CREB NR2B	Ying,et a1,2016 ⁸⁰			
miR-195	Increased	DRG	BDNF	Yu, et a1,2011 ⁷⁷ Shi,et a1,2013 ⁶⁷			







mechanism of changes on the body;

- (2) To identify changes in the epigenetic modification of BDNF in pain, and to observe changes in the DNA, histones, and miRNA of the BDNF gene from the molecular level in the brain region where BDNF expression has changed, in order to understand the effects of these changes on the BDNF gene;
- (3) To clarify the mechanism of epigenetic modification of the BDNF gene in the body under chronic pain, and provide new ideas for the treatment of chronic pain. That is, blocking the pathways or molecules that affect the epigenetic regulation of the BDNF to inhibit changes in the epigenetic modification of the BDNF gene;
- (4) Identification of targets for drug treatment of

related diseases to prevent and treat the occurrence of multiple chronic pain. That is, the use of epigenetically-modified enzyme inhibitors to reverse the changes of the BDNF gene, normalizing the expression of the BDNF gene, thus achieving the purpose of preventing or improving the disease.

References

- Zhong, X.L., et al., Activation of Anterior Cingulate Cortex Extracellular Signal-Regulated Kinase-1 and -2 (ERK1/2) Regulates Acetic Acid-Induced, Pain-Related Anxiety in Adult Female Mice. Acta Histochemica Et Cytochemica, 2012. 45(4): p. 219-225.
- Hecke, O.V., N. Torrance, and B.H. Smith, Chronic pain epidemiology and its clinical relevance. British Journal of Anaesthesia, 2013. 111(1): p. 13.





- Hooten, W.M., Chronic Pain and Mental Health Disorders : Shared Neural Mechanisms, Epidemiology, and Treatment. Mayo Clinic Proceedings, 2016. 91(7): p. 955-970.
- Egger, G., et al., Epigenetics in human disease and prospects for epigenetic therapy. Nature, 2004. 429 (6990): p. 457-463.
- Morgan, H.D., et al., Epigenetic reprogramming in mammals. Human Molecular Genetics, 2005. 14 (1): p. 47-58.
- Feinberg, A.P., Phenotypic plasticity and the epigenetics of human disease. Nature, 2007. 447 (7143): p. 433-440.
- Muotri, A.R. and F.H. Gage, Generation of neuronal variability and complexity. Nature, 2006. 441(7097): p. 1087-93.
- Hammer, P., et al., mRNA-seq with agnostic splice site discovery for nervous system transcriptomics tested in chronic pain. Genome Research, 2016. 20(6): p. 847-860.
- Sun, W., L. Zhang, and R. Li, Overexpression of miR-206 ameliorates chronic constriction injury-induced neuropathic pain in rats via the MEK/ ERK pathway by targeting brain- derived neurotrophic factor. Neuroscience Letters, 2017. 646: p. 68.
- Shin, M.K., H.G. Kim, and K.L. Kim, A novel trimeric peptide, Neuropep-1-stimulating brain-derived neurotrophic factor expression in rat brain improves spatial learning and memory as measured by the Y-maze and Morris water maze. Journal of Neurochemistry, 2011. 116(2): p. 205-216.
- Huang, Y.J., K.H. Lee, and J.W.J.E.N. Grau, Complete spinal cord injury (SCI) transforms how brain derived neurotrophic factor (BDNF) affects nociceptive sensitization. 2017. 288: p. 38-50.
- 12. T, A., et al., Mouse and rat BDNF gene structure and expression revisited. 2010. 85(3): p. 525-535.
- 13. Qing-Rong, L., et al., Rodent BDNF genes, novel promoters, novel splice variants, and regulation by cocaine. 2006. 1067(1): p. 1-12.
- 14. Priit, P., et al., Identification of cis-elements and transcription factors regulating neuronal activity-dependent transcription of human BDNF gene.

2011. 31(9): p. 3295.

- Pruunsild, P., et al., Dissecting the human locus: Bidirectional transcription, complex splicing, and multiple promoters ☆. 2007. 90(3): p. 397-406.
- QR, L., et al., Human brain derived neurotrophic factor (BDNF) genes, splicing patterns, and assessments of associations with substance abuse and Parkinson's Disease. 2010. 134B(1): p. 93-103.
- 17. Tognoli, C., et al., Acute stress alters transcript expression pattern and reduces processing of proBDNF to mature BDNF in Dicentrarchus labrax. 2010. 11(1): p. 1-17.
- Priit, P., et al., Dissecting the human BDNF locus: bidirectional transcription, complex splicing, and multiple promoters. 2007. 90(3): p. 397-406.
- 19. Duric, V. and K.E. McCarson, Effects of analgesic or antidepressant drugs on pain- or stress-evoked hippocampal and spinal neurokinin-1 receptor and brain-derived neurotrophic factor gene expression in the rat. J Pharmacol Exp Ther, 2006. 319(3): p. 1235-43.
- Kerr, B.J., et al., Brain-derived neurotrophic factor modulates nociceptive sensory inputs and NMDA-evoked responses in the rat spinal cord. 1999. 19(12): p. 5138-5148.
- Tang, Y., et al., Interaction between astrocytic colony stimulating factor and its receptor on microglia mediates central sensitization and behavioral hypersensitivity in chronic post ischemic pain model. 2017. 68: p. S0889159117304786.
- 22. Kras, J.V., et al., Brain-derived neurotrophic factor is upregulated in the cervical dorsal root ganglia and spinal cord and contributes to the maintenance of pain from facet joint injury in the rat. 2013. 91(10): p. 1312-1321.
- Fukuoka, T., ., et al., Brain-derived neurotrophic factor increases in the uninjured dorsal root ganglion neurons in selective spinal nerve ligation model. 2001. 21(13): p. 4891.
- 24. Zhu, Z.W., et al., Brain-Derived Neurotrophic Factor (BDNF) Is Upregulated and Associated with Pain in Chronic Pancreatitis. 2001. 46(8): p. 1633-9.
- 25. Sen, S., R. Duman, and G.J.B.P. Sanacora, Serum



brain-derived neurotrophic factor, depression, and antidepressant medications: meta-analyses and implications. 2008. 64(6): p. 527-532.

- Ha, S.O., et al., Expression of brain-derived neurotrophic factor in rat dorsal root ganglia, spinal cord and gracile nuclei in experimental models of neuropathic pain. 2001. 107(2): p. 301-309.
- 27. Zhou, L.J., et al., Limited BDNF contributes to the failure of injury to skin afferents to produce a neuropathic pain condition. 2010. 148(1): p. 148.
- Wang, J., et al., Effects of BDNF on activation of spinal dorsal horn astrocytes in rats with spinal nerve ligation. 2011. 31(11): p. 1578-1582.
- 29. Geng, S.J., et al., Contribution of the spinal cord BDNF to the development of neuropathic pain by activation of the NR2B-containing NMDA receptors in rats with spinal nerve ligation. 2010. 222(2): p. 256-266.
- Coull, J.A.M., et al., BDNF from microglia causes the shift in neuronal anion gradient underlying neuropathic pain. 2005. 438(7070): p. 1017-1021.
- Boulle, F., et al., Epigenetic regulation of the BDNF gene: implications for psychiatric disorders. Molecular Psychiatry, 2012. 17(6): p. 584.
- Martinowich, K., et al., DNA methylation-related chromatin remodeling in activity- dependent BDNF gene regulation. Science, 2003. 302(5646): p. 890-893.
- Ishimaru, N., et al., Differential epigenetic regulation of BDNF and NT-3 genes by trichostatin A and 5-aza-2'-deoxycytidine in Neuro-2a cells. Biochemical & Biophysical Research Communications, 2010. 394(1): p. 173.
- Chao, Y.C., et al., Demethylation regulation of BDNF gene expression in dorsal root ganglion neurons is implicated in opioid-induced pain hypersensitivity in rats. Neurochemistry International, 2016. 97: p. 91-98.
- Ikegame, T., et al., DNA methylation analysis of BDNF gene promoters in peripheral blood cells of schizophrenia patients. Neuroscience Research, 2013. 77(4): p. 208-214.
- Menzies, V., et al., Psychoneuroimmunological relationships in women with fibromyalgia. Biological Research for Nursing, 2013. 15(2): p. 219-225.

- Uchida, H., Y. Matsushita, and H. Ueda, Epigenetic regulation of BDNF expression in the primary sensory neurons after peripheral nerve injury: Implications in the development of neuropathic pain. Neuroscience, 2013. 240(9): p. 147-154.
- Ernest, P., et al., Neuronal activity controls Bdnf expression via Polycomb de-repression and CREB/CBP/ JMJD3 activation in mature neurons. 2016. 7: p. 11081.
- Ernest, P., et al., Neuronal activity controls Bdnf expression via Polycomb de-repression and CREB/CBP/ JMJD3 activation in mature neurons. Nature Communications, 2016. 7: p. 11081.
- Lee, K., et al., Molecular mechanism of Jmjd3-mediated interleukin-6 gene regulation in endothelial cells underlying spinal cord injury. 2012. 122(2): p. 272.
- 41. Cascante, A., et al., Gene-specific methylation control of H3K9 and H3K36 on neurotrophic BDNF versus astroglial GFAP genes by KDM4A/C regulates neural stem cell differentiation. 2014. 426(20): p. 3467-3477.
- Zhou, J., et al., The Histone Demethylase JMJD2A Regulates the Expression of BDNF and Mediates Neuropathic Pain in Mice. Experimental Cell Research, 2017. 361(1).
- 43. Schenkel berger, M., et al., Expression regulation by a methyl-CpG binding domain in an E. coli based, cell-free TX-TL system. Physical Biology, 2017. 14(2).
- 44. Schenkel berger, M., et al., Expression regulation by a methyl-CpG binding domain in an E. coli based, cell-free TX-TL system. 2017. 14(2).
- 45. Fuks, F., et al., The methyl-CpG-binding protein MeCP2 links DNA methylation to histone methylation. Journal of Biological Chemistry, 2003. 278(6): p. 4035-4040.
- Wang, Y., et al., Intrathecal 5-azacytidine inhibits global DNA methylation and methyl - CpG-binding protein 2 expression and alleviates neuropathic pain in rats following chronic constriction injury. Brain Research, 2011. 1418(42): p. 64-69.
- Zhang, Z., et al., MeCP2 Repression of G9a in Regulation of Pain and Morphine Reward. Journal of Neuroscience the Official Journal of the Society for Neuroscience, 2014. 34(27): p. 9076.







- Lubin, F.D., T.L. Roth, and J.D.J.J.o.N.t.O.J.o.t.S.f.N. Sweatt, Epigenetic regulation of BDNF gene transcription in the consolidation of fear memory. 2008. 28(42): p. 10576.
- Tajerian, M., et al., Peripheral Nerve Injury Is Associated with Chronic, Reversible Changes in Global DNA Methylation in the Mouse Prefrontal Cortex. Plos One, 2013. 8(1): p. e55259.
- Uchida, S., et al., Epigenetic status of Gdnf in the ventral striatum determines susceptibility and adaptation to daily stressful events. Neuron, 2011. 69 (2): p. 359-372.
- Ooi, L. and I.C. Wood, Chromatin crosstalk in development and disease: lessons from REST. Nature Reviews Genetics, 2007. 8(7): p. 544.
- Hsieh, M.C., et al., Tet1-dependent epigenetic modification of BDNF expression in dorsal horn neurons mediates neuropathic pain in rats. 2016. 6: p. 37411.
- Uchida, H., Y. Matsushita, and H.J.N. Ueda, Epigenetic regulation of BDNF expression in the primary sensory neurons after peripheral nerve injury: Implications in the development of neuropathic pain. 2013. 240(9): p. 147-154.
- 54. Tao, W., et al., Persistent inflammation-induced up-regulation of brain-derived neurotrophic factor (BDNF) promotes synaptic delivery of alpha-amino-3-hydroxy-5- methyl-4-isoxazolepropionic acid receptor GluA1 subunits in descending pain modulatory circuits. J Biol Chem, 2014. 289(32): p. 22196-204.
- 55. Tao, W., et al., Brainstem brain-derived neurotrophic factor signaling is required for histone deacetylase inhibitor-induced pain relief. 2015. 87(6): p. 1035.
- 56. Kai, L., et al., Epigenetic upregulation of Cdk5 in the dorsal horn contributes to neuropathic pain in rats. 2014. 25(14): p. 1116-1121.
- Wang, X., et al., The etiological changes of acetylation in peripheral nerve injury-induced neuropathic hypersensitivity. Mol Pain, 2018. 14: p. 1744806918798408.
- Ishimaru, N., et al., Differential epigenetic regulation of BDNF and NT-3 genes by trichostatin A and 5-aza-2'-deoxycytidine in Neuro-2a cells. 2010. 394(1):

p. 173-177.

- 59. Su, C.L., et al., Epigenetic regulation of BDNF in the learned helplessness-induced animal model of depression. 2016. 76(1): p. 101-110.
- Li, S., et al., Epigenetic regulation of reelin and brain-derived neurotrophic factor genes in long-term potentiation in rat medial prefrontal cortex. 2012. 97 (4): p. 425-440.
- Uchida, H., Y. Matsushita, and H. Ueda, Epigenetic regulation of BDNF expression in the primary sensory neurons after peripheral nerve injury: implications in the development of neuropathic pain. Neuroscience, 2013. 240: p. 147-54.
- Winston, J.H., Q. Li, and S.K. Sarna, Chronic prenatal stress epigenetically modifies spinal cord BDNF expression to induce sex specific visceral hypersensitivity in offspring. Neurogastroenterology & Motility, 2014. 26(5): p. 715–730.
- Zhu, X., et al., Curcumin alleviates neuropathic pain by inhibiting p300/CBP histone acetyltransferase activity-regulated expression of BDNF and cox-2 in a rat model. PLoS One, 2014. 9(3): p. e91303.
- 64. Genda, Y., et al., microRNA changes in the dorsal horn of the spinal cord of rats with chronic constriction injury: A TaqMan® Low Density Array study. International Journal of Molecular Medicine, 2013. 31 (1): p. 129.
- Mellios, N., et al., Molecular determinants of dysregulated GABAergic gene expression in the prefrontal cortex of subjects with schizophrenia. Schizophrenia Research, 2010. 117(2–3): p. 1006.
- Yu, B., et al., Profile of MicroRNAs following Rat Sciatic Nerve Injury by Deep Sequencing: Implication for Mechanisms of Nerve Regeneration. Plos One, 2011. 6 (9): p. e24612.
- 67. Shi, G., et al., Increased miR-195 aggravates neuropathic pain by inhibiting autophagy following peripheral nerve injury. Glia, 2013. 61(4): p. 504-512.
- Su, M., et al., MeCP2 controls hippocampal brain-derived neurotrophic factor expression via homeostatic interactions with microRNA- 132 in rats with depression. Molecular Medicine Reports, 2015. 12 (4): p. 5399.





- Kynast, K.L., et al., Modulation of central nervous system-specific microRNA-124a alters the inflammatory response in the formalin test in mice. Pain, 2013. 154 (3): p. 368-376.
- Willemen, H.L., et al., MicroRNA-124 as a novel treatment for persistent hyperalgesia. Journal of Neuroinflammation,9,1(2012-06-25), 2012. 9(1): p. 143.
- Lin, C.R., et al., Intrathecal miR-183 delivery suppresses mechanical allodynia in mononeuropathic rats. Eur J Neurosci, 2014. 39(10): p. 1682-9.
- 72. Neumann, E., et al., Expression changes of microRNA-1 and its targets Connexin 43 and brain-derived neurotrophic factor in the peripheral nervous system of chronic neuropathic rats. Mol Pain, 2015. 11: p. 39.
- Lin, C.R., et al., Intrathecal miR 183 delivery suppresses mechanical allodynia in mononeuropathic rats. European Journal of Neuroscience, 2014. 39(10): p. 1682-1689.
- 74. Sun, W., L. Zhang, and R. Li, Overexpression of miR-206 ameliorates chronic constriction injury-induced neuropathic pain in rats via the MEK/ERK pathway by targeting brain- derived neurotrophic factor. Neurosci Lett, 2017. 646: p. 68-74.
- 75. Li, H., et al., Upregulation of miR-375 level ameliorates morphine analgesic tolerance in mouse dorsal root ganglia by inhibiting the JAK2/STAT3 pathway. Journal of Pain Research, 2017. 10: p. 1279.
- 76. Hu, X.M., et al., Downregulation of miR-219 enhances brain-derived neurotrophic factor production in mouse dorsal root ganglia to mediate morphine analgesic tolerance by upregulating CaMKIIγ. Molecular Pain,12, (2016-9-01), 2016. 12(47): p. 14670-14683.
- 77. Yu, B., et al., Profile of microRNAs following rat sciatic nerve injury by deep sequencing: implication for mechanisms of nerve regeneration. PLoS One, 2011. 6 (9): p. e24612.
- 78. Guodong, S., et al., Increased miR-195 aggravates neuropathic pain by inhibiting autophagy following peripheral nerve injury. 2013. 61(4): p. 504-512.
- 79. Mellios, N., et al., Molecular determinants of dysregulated GABAergic gene expression in the prefrontal cortex of subjects with schizophrenia. Biol

Psychiatry, 2009. 65(12): p. 1006-14.

- Ying, L., et al., CREB-regulated transcription coactivator 1 enhances CREB-dependent gene expression in spinal cord to maintain the bone cancer pain in mice. Mol Pain., 2016. 12.
- Li, X., Y. Zhang, and T.J.J.o.M.N. Dong, Inhibition of MicroRNA-221 Alleviates Neuropathic Pain Through Targeting Suppressor of Cytokine Signaling 1. 2016. 59 (3): p. 1-10.